

Delivery of SERCA2a as a Treatment for Heart Failure

A number of cardiac abnormalities are associated with advanced heart failure (HF), all leading to a reduced ability of the heart to pump blood and supply adequate oxygenated blood to the body. Reduced contractility of the individual cardiomyocytes is the main factor resulting in reduced global cardiac output, and intracellular calcium (Ca^{2+}) flux is central to this cellular abnormality.

Regulation of excitation-contraction coupling in cardiomyocytes is driven by Ca^{2+} flux between the sarcoplasmic reticulum (SR) and the cytosol. Electrical excitation of the sarcolemma leads to the opening of voltage gated L-type Ca^{2+} channels, allowing the entry of a small amount of Ca^{2+} into the cell (Figure below). This influx leads to opening of the ryanodine receptors located on the SR membrane and a large amount of Ca^{2+} is released from the SR into the cytosol. The resulting increase in cytosolic Ca^{2+} activates the myofilaments resulting in contraction. [1, 2] During relaxation, Ca^{2+} is pumped back into the SR by the SR Ca^{2+} ATPase (SERCA2a) pump and extruded extracellularly by the sarcolemmal $\text{Na}^+/\text{Ca}^{2+}$ exchanger. The contribution of SERCA2a in humans for removing intracellular calcium is ~75% and ~25% for the $\text{Na}^+/\text{Ca}^{2+}$ exchanger. The Ca^{2+} pumping activity of SERCA2a is regulated by phospholamban. In the unphosphorylated state, phospholamban inhibits the Ca^{2+} -ATPase, whereas phosphorylation of phospholamban by cAMP-dependent protein kinase and by Ca^{2+} -calmodulin dependent protein kinase releases the inhibition.

In summary, in the mammalian heart, the intracellular Ca^{2+} movements are tightly regulated at various levels within the cell. SERCA2a plays a central role in regulating contraction and relaxation by its role in controlling the level of Ca^{2+} in the cytosol and SR in the cardiomyocyte. [3]

Defects in Excitation-Contraction coupling, due to abnormal expression and/or function of calcium handling and transport proteins, are a hallmark of cardiac dysfunction. These defects manifest in changes in the calcium transient: reduced amplitude, increased duration, and prolonged decay - the consequence of which is decreased contractility and reduced cardiac output. Regardless of the etiology of heart failure, the role of reduced SR calcium load has been well established in terms decreased SR calcium uptake, decreased SR calcium content, and calcium leak from the SR. [4] [5] This is primarily due to a decrease in SR calcium uptake because of SERCA2a dysfunction. In both animal models of and patients with heart failure, there is a reduction in SERCA2a expression (mRNA and protein levels) and activity. [6]

Increasing the levels of SERCA2a protein in cardiomyocytes has been shown in human cells, experimental rodent and large animal models to normalize the abnormally high diastolic levels of cytosolic calcium typical of HF and improve contractile and clinical outcomes, as described below. [7-9] [10-13]

SARDOCOR proposes to investigate gene transfer as a method to restore SERCA2a function in heart failure patients using a recombinant adeno-associated viral vector (AAV), which consists of an AAV serotype 1 capsid and contains the human SERCA2a cDNA flanked by ITRs derived from AAV serotype 2 (AAV1/SERCA2a).

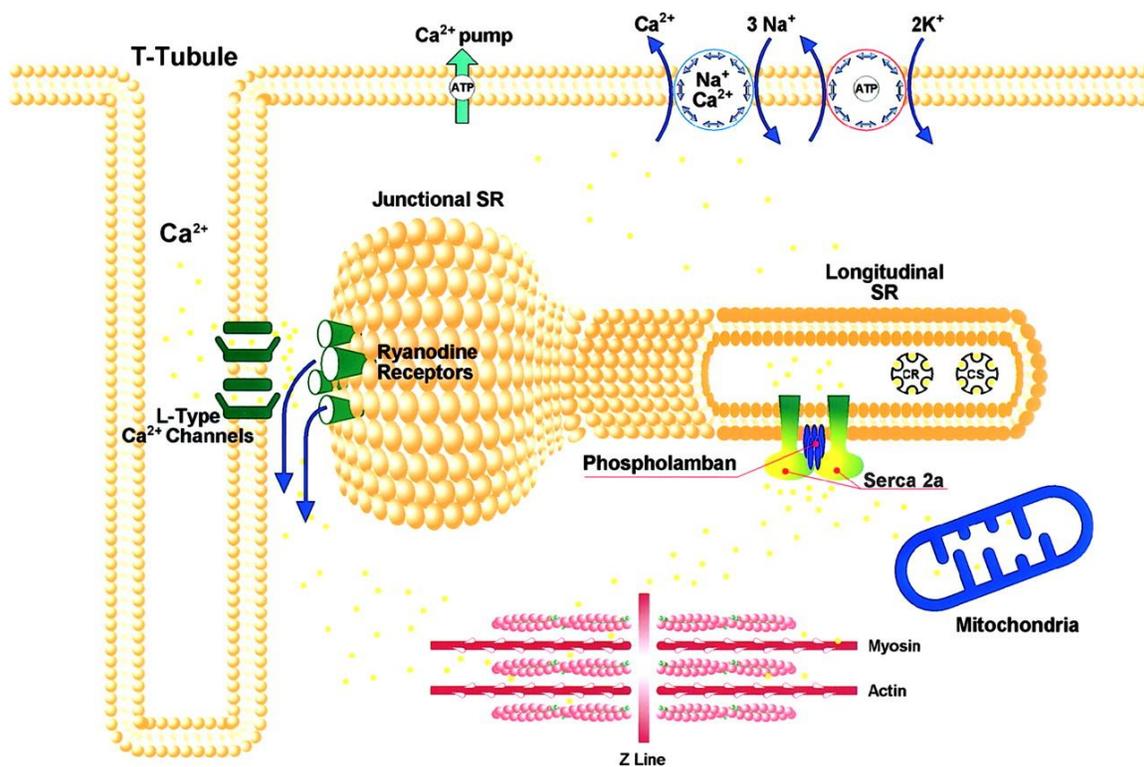


Figure : Ca²⁺ Handling Proteins Involved in Intracellular Ca²⁺ Movement

In addition to the role in cardiomyocytes, SERCA2a also has a role in skeletal muscle, the diaphragm, vascular smooth muscle, and endothelial cells. Both moderate and high intensity exercise increase the relative SERCA2a isoform expression in cardiac and skeletal muscle cells. SERCA2a gene transfer also results in increased coronary blood flow in a diabetic rat model. Finally, increasing SERCA2a levels via gene transfer inhibits vascular smooth muscle proliferation.

References:

1. [Bers, D., *Cardiac excitation-contraction coupling*. Nature, 2002. 415: p. 198-205.](#)
2. [Bers, D., *Calcium fluxes involved in control of cardiac myocyte contraction*. Circulation Research, 2000. 87\(4\): p. 275-81.](#)
3. [Bers, D., *Sarcoplasmic reticulum Ca release in intact ventricular myocytes*. Frontiers in Bioscience, 2002. 7: p. d1697-711.](#)
4. [Kranias, E.G. and R.J. Hajjar, *Modulation of cardiac contractility by the phospholamban/SERCA2a regulatome*. Circ Res, 2012. 110\(12\): p. 1646-60.](#)
5. [Gorski, P.A., D.K. Ceholski, and R.J. Hajjar, *Altered myocardial calcium cycling and energetics in heart failure--a rational approach for disease treatment*. Cell Metab, 2015. 21\(2\): p. 183-194.](#)
6. [Meyer, M., et al., *Alterations of sarcoplasmic reticulum proteins in failing human dilated cardiomyopathy*. Circulation, 1995. 92\(4\): p. 778-84.](#)
7. [del Monte, F., et al., *Restoration of contractile function in isolated cardiomyocytes from failing human hearts by gene transfer of SERCA2a*. Circulation, 1999. 100\(23\): p. 2308-11.](#)
8. [del Monte, F., et al., *Improvement in survival and cardiac metabolism after gene transfer of sarcoplasmic reticulum Ca\(2+\)-ATPase in a rat model of heart failure*. Circulation, 2001. 104\(12\): p. 1424-9.](#)
9. [Miyamoto, M.I., et al., *Adenoviral gene transfer of SERCA2a improves left-ventricular function in aortic-banded rats in transition to heart failure*. Proc Natl Acad Sci U S A, 2000. 97\(2\): p. 793-8.](#)
10. [Beeri, R., et al., *Gene delivery of sarcoplasmic reticulum calcium ATPase inhibits ventricular remodeling in ischemic mitral regurgitation*. Circ Heart Fail, 2010. 3\(5\): p. 627-34.](#)
11. [Kawase, Y., et al., *Reversal of cardiac dysfunction after long-term expression of SERCA2a by gene transfer in a pre-clinical model of heart failure*. J Am Coll Cardiol, 2008. 51\(11\): p. 1112-9.](#)
12. [Prunier, F., et al., *Prevention of ventricular arrhythmias with sarcoplasmic reticulum Ca²⁺ ATPase pump overexpression in a porcine model of ischemia reperfusion*. Circulation, 2008. 118\(6\): p. 614-24.](#)
13. [Sakata, S., et al., *Restoration of mechanical and energetic function in failing aortic-banded rat hearts by gene transfer of calcium cycling proteins*. J Mol Cell Cardiol, 2007. 42\(4\): p. 852-61.](#)